

Bioformulation in Biological Control for Plant Diseases - A Review

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Abstract

Bioformulation for plant growth promotion continue to inspire research and development in many fields. Increase in soil fertility, plant growth promotion, and suppression of phytopathogens are the targets of the bioformulation industry that leads to the development of ecofriendly environment. The rhizosphere bacteria have immense application in sustainable agriculture as ecofriendly biofertilizer and biopesticides. Intensive commercial farming involves excessive use of chemical fertilizers and pesticides. It is feared that practice of using chemical fertilizers and pesticides continually would result in gradual aggravation of soil fertility. The aim of the review is to assess the biological control of plant pathogens in with the effective development of bioinoculant industry.

Keywords: *carrier based bioformulation, growth promotion, biocontrol, biofertilizer, rhizosphere bacteria, plant pathogens.*

I. INTRODUCTION

The development and use of *Trichoderma* and *Talaromyces* -based bioformulations with talc and rice bran may be a potentially effective method in controlling *Rhizoctonia solani* induced sugar beet damping-off disease. May have practical application in the formulation of disease management strategies in an integrated pest management (IPM) program in which reduction of yield loss, reduction of chemical pesticide application and protection of the agricultural environment and biological resources are the main concerns (Kakvan et al, 2013).

Trichoderma species have shown highly promising results against *Fusarium oxysporum* f. sp. *melonis* under different experimental conditions. In the studies of Suárez-Estrella et al. (2007), *Trichoderma harzianum* 2413 reduced the incidence of melon wilt in a greenhouse experiment. In previous research, we showed that selected isolates of *Trichoderma* spp. were efficient to control soil-borne pathogens of melon in field conditions (Gava and Menezes, 2012). *Trichoderma* spp. are the most widely studied biological control agents (BCAs) for root and shoot pathogens, applied even in post-harvest (Woo et al., 2014). *Trichoderma* species are soil inhabitants,

competitive saprophytes, and facultative mycoparasites that can colonize the soil and rhizosphere (Harman et al., 2004). Their mechanisms of action include the lysis of fungal hyphae with enzymes such as chitinase, proteases, and glucanases (Shahid et al., 2014), the induction of phytoalexin accumulation by the host (Yedidia et al., 2003), the production of antibiotics (El-Hasan et al., 2006; Reino et al., 2008) and modulation of plant hormones (Martínez-Medina et al., 2010).

Using bioformulations with entomopathogen fungi for biological control of insect pests we can point out the easiness of production of its infective units on a commercial scale, the simplicity of usage in field conditions, the low cost of its utilization, and mainly, the reduction on environmental impact (Lopes et al, 2013, Nussenbaum et al, 2013, Borisade and Magan, 2014, Blanford et al, 2011, Zahran et al, 2013). Interactions between entomopathogenic fungi with phytosanitary products, such as chemical insecticides (e.g. Decis OC), fungicides (e.g. Manzate 800) or herbicides (e.g. Granoxone) are important to evaluate new formulations, since it can be positive when an additive or synergistic action occurs with the entomopathogen and the product. The formulation is a need for compatibility testing, seeking more selective products and able to promote the conservation of the pathogen in the field for a longer period of time (Borges et al, 2011; Vidau et al, 2011).

In India, specific formulations of *Pseudomonas* spp. strains have been developed to combat various pathogens in rice, banana and pigeon pea (Rabindran & Vidhyasekaran 1996; Vidhyasekaran et al. 1997a, 1997b). With a view to standardize specific *Pseudomonas* spp. strains for the management of red rot disease of sugarcane, we have optimized talc formulations for sett treatment and soil application and efficacy of different strains against the disease development in the cane stalks.

Biological control using microbial antagonists has been shown to be a suitable and ecologically friendly candidate for the replacement of chemical pesticides. Different bacterial and fungal antagonists have proved to be potential biocontrol agents against many plant pathogenic fungi and bacteria (Dutta, 1981;

Papavizas, 1995; Matta and Garibadli, 1997; Menendez and Godeas, 1998; Adebajo and Bankole, 2004; Eziashi et al., 2006; Naraghi et al., 2006, 2007; Gentili et al., 2008; Jahanifar et al., 2008; Heydari and Pessarakli, 2010; Gerami et al., 2013). The use of chemical fungicides as seed treatment is the most common strategy for controlling this disease in the field which most of the time is not effective due to the long term application and appearance of resistant races of the pathogen (Weller, 1991; Heydari and Pessarakli, 2010). Several microorganisms have been reported as plant pathogen antagonists, but only a small number were applied on a commercial scale. Most of this is due to a lack of consistency of the results from field trials (Fravel, 2005).

The high doses of chemical fungicides can result in environmental pollution, deterioration of human health, and increase in resistance of the target fungi to fungicides. Biological control has been investigated for its potential to provide a more viable and sustainable means to control southern stem rot. Attempts to develop biological control of *Sclerotium rolfsii* have been reported in many crops such as peanut and soybean (Ambang et al., 2008; Ozgonen et al., 2010; Ika et al., 2011; Rakh et al., 2011).

Plant Growth Promontory Bioformulation

PGPR would be beneficial not only for plant growth but also for reduction of insect pest attack. One of the common means of application of bacterial inoculants to soil is in the form of bioformulations. Viability of inoculum in an appropriate formulation for a certain length of time is important for commercialization of the technology (Bashan, 1998). Previous reports are also available where *Bacillus* bioformulations could survive upto one year (El-Hassan and Gowen, 2006). Carrier based preparations of two PGPR such as *Bacillus subtilis* and *Pseudomonas corrugata* developed in formulations were also evaluated for their growth promotion, rhizosphere colonization and viability under storage (Trivedi et al, 2005).

Viswanathan and Samiyappan (2008) reported that the potential of suppressing *Colletotrichum falcatum* causing systemic infection in sugarcane stalks by *Pseudomonas* spp. strains under field conditions. These studies also proved that *Pseudomonas* spp. treatment has improved cane yield and sugar yield in the trials. Different sugar mills in the country have evinced keen interest in promoting this new technique for the management of *C. falcatum* in sugarcane.

Chakravarty and Kalita (2011) showed the suppression of bacterial wilt with concomitant

improvement in yield and yield attributes of bio formulation treated crops compared to inoculated control reinforces *Pseudomonas fluorescens* as a biocontrol agent of bacterial wilt in brinjal as well as plant growth promoting rhizobacteria (Ramesh, Joshi and Ghanekar 2008). However, intensive screening of indigenous strains of *P. fluorescens*, development of improved carriers and large scale field trials under different climatic conditions are necessitated for evolving formulations with better disease control activity in the field.

Raj et al, Chuaboon and Prathuangwong (2003, 2007) reported to indicate a future possibility that plant growth promoting rhizobacteria bioformulations can be used to promote growth and health of economic crops.

A bioformulation using humic acid and a suitable microorganism namely, *Pseudomonas fluorescens* has been developed to replace chemical fertilizers. This liquid formulation in addition to facilitates long shelf life, zero contamination, no need of carriers, convenience of handling, storage and transportation has easy to use with irrigation. The mixed formulation of humic acid along with the microorganism namely *Pseudomonas fluorescens* can be used for the dual purpose viz., crop protection and enhanced production. The liquid bio-formulation was tested and compared for viability as well as its inhibitory characteristics against *Fusarium oxysporum*, a fungus which cause wilt of tomato. Field studies were conducted for two crop varieties- radish and tomato (Agrawal Pushpa et al, 2013).

Chakraborty et al (2013) have shown that all three isolates have potential as plant growth promoters to increase the growth of tea plants in experimental plot. Increase in growth was associated with phosphate solubilization, defense enzymes as well as increased accumulation of phenolics. Viabilities of the isolates in bioformulations of talc, saw dust and rice husk were also examined. In comparison to *Serratia marcescens*, bioformulations of *Bacillus amyloliquefaciens* and *B. pumilus* were more useful in field application due to the formation of endospores by bacilli.

Biological control in bioformulation

Biological control of plant pathogen is becoming an important component of plant disease management practices. This alternative control strategy can solve many persistent problems in agriculture including fungicide residues causing environmental pollution and human health hazard, and also inducing pathogen resistance (Commare et al., 2002; Cook, 2002;

Bharathi et al., 2004; Chaluvvaraju et al., 2004; Anitha and Rabeeth, 2009; Chen et al., 2009; Ardakani et al., 2010,2011; Haggag and Wafaa, 2012).The integrated management of cacao black pod disease, biological control was first initiated in Cameroon early in the 2000's and has led to the development of a formulation using mycoparasitic strains of *Trichoderma asperellum* as active ingredient. Field applications showed that the formulation improved the flowering activity of cacao trees as well as pod development, while reducing *Phytophthora* infection in flower cushions (Tondje et al., 2007; Deberdt et al., 2008).

A formulated product must be economical to produce, easy to apply in the crop production system, efficacies with an adequate number of viable cells when used, and a shelf-stable formulated product retraining biocontrol activity comparable to fresh cells of the agent. Delivery systems employing biocontrol agent include dust or powder, alginate pellet, and starch or extruded granule that the effective strains are necessary to be grown in various organic and inert carriers, such as diatomaceous earth, manure or animal dung (Raj et al., 2003; Schisler et al., 2004; Sharathchandra et al., 2004; Amran, 2006; Pushpalatha et al., 2007; Preecha and Prathuangwong, 2009; Omer, 2010; Senthilraja et al., 2010; Siripornvisal and Trilux, 2011). The yeast *Meyerozyma caribbica* L6A2 is an efficient biocontrol for the phytopathogenic *Colletotrichum gloeosporioides*, presenting different antagonistic mechanisms of action such as competition for space and nutrients, production of hydrolytic enzymes, parasitism and biofilm formation through quorum sensing. According to the action mechanisms observed, it is assessed that the presence of yeast cells is necessary in the formula to carrying out biological control. This information needs to be taken account for further studies, especially in formulation and large scale production. (Bautista-Rosales et al. 2013).

This investigation is aimed at providing a better biocontrol action against fungal pathogens as well as insects affecting maize crop. Nanosilica treated maize possesses rough leaf surface and good physical strength than that treated with bulk silica due to higher silica accumulation. A combination of *Pseudomonas fluorescence* and silica nanoparticles in soil enhances phenolic activity and hence reduces the stress by the suppression of responsive enzymes in maize. This elevated level of phenols is found to induce silica accumulation in leaf epidermis, thereby conferring a protective physical barrier as well as induced disease resistance. Thus, one can formulate an effective biofertilizer/biocomposite for sustainable crop cultivation by using nanosilica (Suriyaprabha Rangaraj et al, 2014).

Damam et al. (2015) suggested that simultaneous screening of rhizobacteria for growth and yield promotion under pot and field experiment is a good tool to select effective PGPR strains (*Pantoea agglomerans* (Cf 7), *Pseudomonas putida* (Te 1), *Bacillus subtilis* (Cf 60) and *Pseudomonas* sp. (Av 30) for biofertilizer development biotechnology and potential biocontrol agent against pathogen (*Macrophomina phaseolina*) individually and also in combinations. (Damam et al. 2015).

The biological control of pathogens, although subject to numerous ecological limitations, is expected to become an important part of the control measures employed against *Ralstonia solanacearum*. In the present study, the application of *Trichoderma parareesei* + *Trichoderma parareesei* + *Pseudomonas fluorescens* + *Bacillus subtilis* + *Azotobacter chroococcum* based bio-formulations promises as an effective biocontrol option along with better plant growth, yield and soil health management of Tomato plant (Bharat et al. 2016).

Research Areas for Development of Bioformulation

Biocontrol approaches may help to develop bioformulations an eco-friendly control strategy for managing plant disease (Heydari and Misaghi, 1998, 2003; Bharathi et al., 2004; Heydari and Gharedaghli, 2007 ;).

The development of stable formulations of antagonistic bacteria and other biocontrol agents is of great importance to many countries, especially those where subsistence agriculture is prominent, soil-borne diseases are the main problem, and fungicides are unaffordable. Formulation and establishment of biocontrol agents are very important for their effectiveness. The formulations we have developed and tested can be used for controlling Plant diseases and possibly other plant-pathogen combinations. They have the potential to replace chemical fungicides and to be utilized as an important component of integrated pest management (IPM), which is a promising approach to a sustainable agriculture (Ardakani et al, 2010).

A bioformulation can improve product stability and also protect bacteria against different environmental conditions and provide initial food source. Application of PGPR either to increase crop health or to manage plant diseases depends on the development of commercial formulations with suitable carriers that support the survival of bacteria for a considerable length of time., it is imperative to evaluate the survival of the immobilized bacteria in different

carriers and also their ability to retain attributes for plant growth promotion (Aeron et al., 2011).

Shelf Life Increased for Bioformulation

Research on nitrogen fixation and phosphate solubilization by plant growth promoting rhizobacteria(PGPR) is progress on but little research can be done on potassium solubilization which is third major essential macronutrient for plant growth. This will not only increase the field of the inoculants but also create confidence among the farmers for their use. A part from that future research in optimizing growth condition and increased shelf life of PGPR products, not phytotoxic to crop plants, tolerate adverse environmental condition, higher yield and cost effective PGPR products for use of agricultural farmer will be also helpful (Gupta et al. 2015).

The talc based bioformulation of *Pseudomonas fluorescens* RRb-11 isolate showed maximum shelf life and survivability in rhizosphere to reduce disease intensity of bacterial blight of rice and thereby increase yield when applied as seed treatment, seedling root dip and soil drenching in combination (Jambhulkar and Sharma, 2014).

The most common solutions to this problem of survival time have been air-dried and lyophilized preparations (Kosanke et al. 1992). The lowered water

content in the final product is responsible for long-term survival during storage. The bacteria in the formulation remain inactive, resistant to environmental stresses, insensitive to contamination, and are more compatible with fertilizer application (Bashan 1998). The dehydration phase is perhaps the most critical of the entire formulation process especially for nonspore-forming bacteria (Shah-Smith and Burns 1997).

Bacterial survival is affected by several variables: the culture medium used for bacterial cultivation, the physiological state of the bacteria when harvested from the medium, the use of protective materials, the type of drying technology used, and the rate of dehydration (Paul et al. 1993).

One of the major challenges for the inoculant industry is to develop an improved formulation that provides high shelf-life, high number of viable cells, protection against soil environment, convenience to use, and cost effective (Smith 1992). More studies on the practical aspects of mass-production and formulation need to be undertaken to make new bioformulations that are stable, effective, safer, and more cost-effective. There is an urgent need to develop a definite correlation between agriculturists, microbiologists, biotechnologists, industrialists, and farmers (Fig. 1).

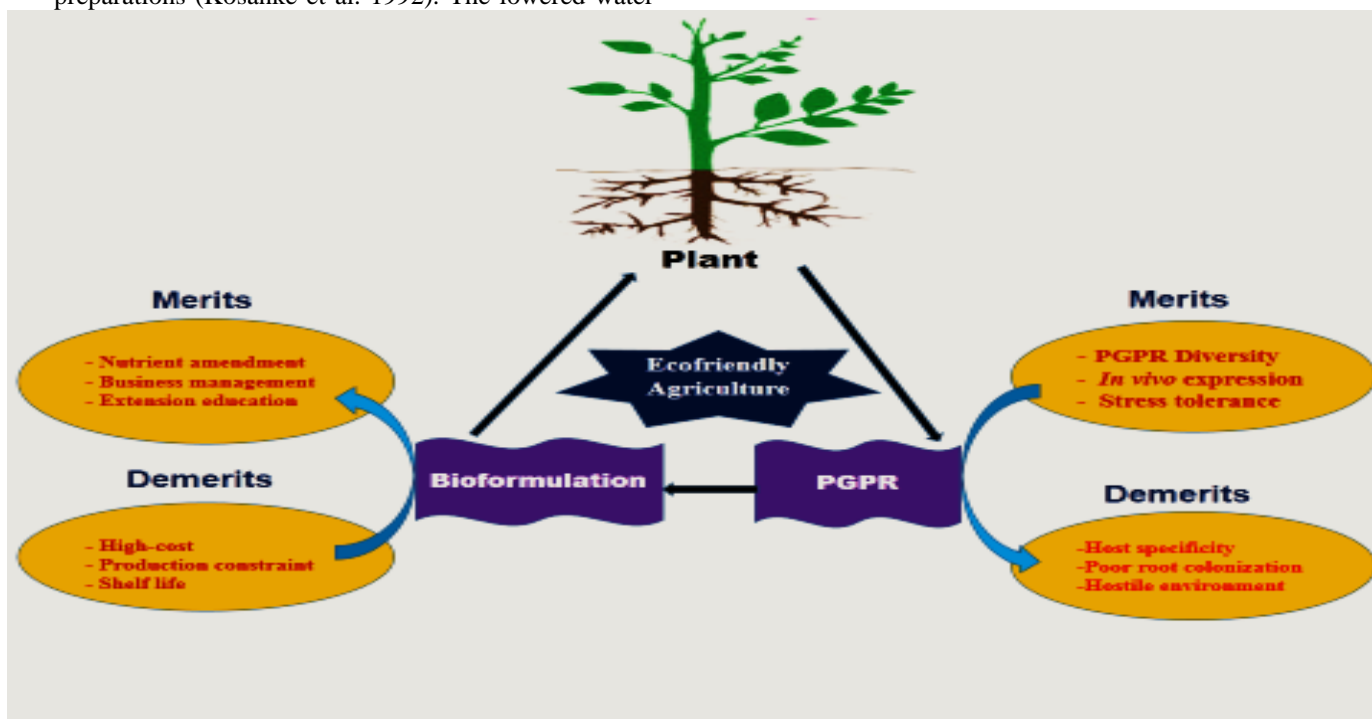


Fig. 1 Research and Development Strategies for Bioformulation Technology

Bioformulation Strategy

The results of the present study in the development of some new bioformulations are promising and may have practical application in the biological promotion of the growth of garlic in the field conditions. Obtaining positive results in the field conditions may replace or reduce the application of harmful chemical fertilizers and lead the garlic growers to increase the yield and production of this important crop and protect the agricultural environment and biological resources (Razak et al. 2015).

Peat and soil rich in organic matter are generally used in the preparation of legume inoculants and constitute a suitable carrier for the purpose. Peat and lignite, though good carriers, are not easily available and are expensive. The low cost and easily availability of carrier material are the major requirements for bioformulations in developing countries (Saha et al. 2001).

The carbon sources and minerals have been shown to have an important role in antifungal metabolite production by *Pseudomonas* biocontrol agents, suggesting that nutrient amendments to formulations may also be a useful strategy for improving biocontrol efficacy (Duffy and De'fago 1999). Soil amendment with chitin showed increase of the chitinolytic microbial populations and significantly reduced the incidence of fungal diseases in celery (Bell et al. 1998). Chitin supplementation supports the survival of *Bacillus cereus* and *Bacillus circulans* in the groundnut phylloplane and resulted in better control of early and late leaf spot disease (Kishore et al. 2005).

Remarkably low percentage of endospore formers was observed that survived after drying (Validov et al. 2007). This has been termed anhydrobiotic engineering (Fages 1992), in reference to anhydrobiotic organisms which naturally exhibit extreme desiccation tolerance (Validov et al. 2009). Similar observations of Garcí ´ a de Castro et al. (2000) demonstrate the potential of this novel biotechnology for stabilizing nonsporulating organisms.

Reasons for Doing the Study

The use of biopesticides include increased environmental awareness, the pollution potential and health hazards from many conventional pesticides, as well as increasing global demand for organically grown food. Biopesticides can be used in rotation with conventional pesticides when used in Integrated Pest Management (IPM) programs. Such programs can offer high crop yields while dramatically reducing the use of conventional pesticides.

Conclusion

The survey of both conventional and organic growers indicates an interest in using biological products (Rzewnicki 2000), suggesting that the market potential of bioformulations will increase in coming years. Internationally, organic sales grew by 8% in 2010 and sales are now valued at €44.5 billion. Strong growth has continued in all the major European markets, and the US, and the outlook for this year is positive. The organic market in China has quadrupled in the past five years, while Organics Brazil reports an annual growth rate of 40% in the Brazilian market. Market analysts predict that organic sales in Asia will grow by 20% a year over the next three years. Thirty-seven million hectares of land worldwide are now farmed organically. (Ref: www.soilassociationscotland.org).

The global pesticide market was valued at approximately \$40 billion in 2008. This figure increased to nearly \$43 billion in 2009 and is expected to grow at a compound annual growth rate (CAGR) of 3.6% to reach \$51 billion in 2014. BCC Research projects that the global biopesticide and synthetic pesticide market will grow from \$54.8 billion in 2013 to nearly \$61.8 billion by 2014 and to \$83.7 billion by 2019 at a five-year compound annual growth rate (CAGR) of 6.3%, from 2014 through 2019. Biopesticides represent only 2.89% (as on 2005) of the overall pesticide market in India and is expected to increase drastically in coming years. (Ref: <http://www.prnewswire.com/news-releases/global-markets-for-biopesticides-279175451>).

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REFERENCES

- [1] Dutta, B.K., 1981. Studies on some fungi isolated from the rhizosphere of tomato plants and the consequent prospect for the control of Verticillium wilt. *Plant Soil* 63, 209-216.
- [2] Papavizas, G.O., 1995. Trichoderma biology, ecology and potential for biocontrol. *Ann. Rev. Phytopathol* 23, 28-54.
- [3] Matta, A., Garibadi, A., 1997. Control of Verticillium wilt of tomato by pre-inoculation with avirulent fungi. *Eur. J. Plant Pathol.* 83, 457-462.
- [4] Menendez, A.B., Godeas, A., 1998. Biological control of *Sclerotinia sclerotiorum* attacking soybean plants: degradation of the cell wall of this pathogen by *Trichoderma harzianum*. *Mycopathology* 142, 153-160.
- [5] Adebajo, A., Bankole, S.A., 2004. Evaluation of some fungi and bacteria for biocontrol of anthracnose disease of cowpea. *J. Basic Microbiol.* 44, 3-9.
- [6] Eziashi, E.L., Uma, N.U., Adekunle, A.A., Ariede, C.E., 2006. Effect of metabolites produced by *Trichoderma* species

- against *Ceratocystis paradoxa* in culture medium. *Afr. J. Biotechnol.* 5, 703-706.
- [7] Naraghi, L., Heydari, A., Ershad, D., 2006. Sporulation and survival of *Talaromyces flavus* on different plant material residues for biological control of cotton wilt caused by *Verticillium dahlia*. *Iran J. Plant* 42, 381-397.
- [8] Naraghi, L., Zareh-Maivan, H., Heydari, A., Afshari-Azad, H., 2007. Investigation of the effect of heating, vesicular arbuscular mycorrhiza and thermophilic fungus on cotton wilt disease. *Pak J. Biol. Sci.* 10, 1596-1603.
- [9] Gentili, A., Mariotti, E., Vincenzi, A., Mazzaglia, A., Heydari, A., Schaad, N.W., Varvaro, L., Balestra, G.M., 2008. Dieback (Moria) of hazelnut: isolation and characterization of two potential biocontrol agents. *J. Plant Pathol.* 90,383-386.
- [10] Jahanifar, H., Heydari, A., Hassanzadeh, N., Zamanizadeh, H.R., Rezaee, S., Naraghi, L., 2008. A comparison between antibiotic-resistant mutants of antagonistic bacteria and their wild types in biological control of cotton seedling damping-off disease. *J. Biol. Sci.* 8, 914- 919.
- [11] Heydari, A., Pessarakli, M., 2010. A review on biological control of fungal plant pathogens using microbial antagonists. *J. Biol. Sci.* 10, 273-290.
- [12] Gerami, E., Hassanzadeh, N., Abdollahi, H., Ghasemi, A., Heydari, A., 2013. Evaluation of some bacterial antagonists for biological control of fire blight disease. *J. Plant Pathol.* 95, 127- 134.
- [13] Weller, D.M., 1991. Biological control of soil-born plant pathogens in the rhizosphere with bacteria. *Ann. Rev. Phytopathology.* 26, 379-407.
- [14] Ambang, Z., Ndongo, B., Bime, Ngoh, D., Maho, Y., Ntsomboh, G., 2008. Effect of mycorrhizal inoculum and urea fertilizer on diseases development and yield of groundnut crops (*Arachis hypogaea* L.). *Afr. J. Biotechnol.* 7 (16), 2823-2827.
- [15] Ozgonen, H., Akgul, D.S., Erkilic, A., 2010. The effects of arbuscular mycorrhizal fungi on yield and stem rot caused by *Sclerotium rolfsii* Sacc. in peanut. *Afr. J. Agric.Res.* 5 (2), 128-132.
- [16] Ika, R.S., Syamsuddin, D., Nasir, S., Anton, M., 2011. Control of damping off disease caused by *Sclerotium rolfsii* Sacc. using actinomycetes and VAM fungi on soy-bean in the dry land based on microorganism diversity of rhizosphere zone. *Agrivita* 33 (1), 40-46.
- [17] Rakh, R.R., Raut, L.S., Dalvi, S.M., Manwar, A.V., 2011. Biological control of *Sclerotium rolfsii*, causing stem rot of groundnut by *Pseudomonas cf. monteilii* 9. *Rec. Res.Sci. Tech.* 3, 26 -34.
- [18] Lopes RB, Martins I, Souza DA, Faria M. Influence of some parameters on the germination assessment of mycopesticides. *J Invertebr Pathol.* 2013; 112(3):236-242.
- [19] Nussenbaum AL, Lewylle MA, Lecuona RE. Germination, Radial Growth and Virulence to Boll Weevil of Entomopathogenic Fungi at Different Temperatures. *World Appl Sci J.* 2013; 25(8):1134-1140.
- [20] Borisade OA, Magan N. Growth and sporulation of entomopathogenic *Beauveria bassiana*, *Metarhizium anisopliae*, *Isaria farinosa* and *Isaria fumosorosea* strains in relation to water activity and temperature interactions. *Biocontrol Sci Technol.* 2014; 24(9):999-1011.
- [21] Blanford S, Shi W, Christian R, Marden JH, Koekemoer LL, Brooke BD, et al. Lethal and pre-Lethal effects of a fungal biopesticide contribute to substantial and rapid control of malaria Vectors. *Plos One.* 2011; 6(8):e23591.
- [22] Zahran HEDM, Kawanna MA, Bosly HA. Larvicidal Activity and Joint Action Toxicity of Certain Combating Agents on *Culex pipiens* L. Mosquitoes. *Ann Rev Res Biol.* 2013; 3(4):1055-106.
- [23] Borges LR, Vila Nova MX. Association of chemical insecticides and entomopathogenic fungi in Integrated Pest Management – a review. *Ambiência.* 2011; 7(1):179–190.
- [24] Vidau C, Diogon M, Aufauvre J, Fontbonne R, Viguès B, Brunet JL, et al. Exposure to sublethal doses of Fipronil and thiacloprid highly increases mortality of honeybees previously infected by *Nosema ceranae*. *Plos One.* 2011; 6(6):e21550.
- [25] Rabindran R, Vidhyasekharan P. 1996. Development of a formulation of *Pseudomonas fluorescens* PfALR2 for management of rice sheath blight. *Crop Protection* 15:715–721.
- [26] Vidhyasekaran P, Sethuraman K, Rajappan K, Vasumathi K. 1997a. Powder formulations of *Pseudomonas fluorescens* to control pigeon pea wilt. *Biological Control* 8:166–171.
- [27] Vidhyasekaran P, Rabindran R, Muthamilan M, Nayyar K, Rajappan K, Subramanian N, Vasumathi K. 1997b. Development of powder formulation of *Pseudomonas fluorescens* for control of rice blast. *Plant Pathology* 46: 291–297.
- [28] R. Viswanathan and R. Samiyappan (2008) Bio-formulation of fluorescent *Pseudomonas* spp. induces systemic resistance against red rot disease and enhances commercial sugar yield in sugarcane. *Archives Of Phytopathology And Plant Protection*, 41:5, 377-388.
- [29] G.Chakravarty and M.C.Kalita (2011) Management of bacterial wilt of brinjal by *Pseudomonas fluorescens* based bioformulation. *ARN Journal of Agricultural and Biological Science.* 6(3), 1-11.
- [30] Ramesh R, Joshi A.A and Ghanekar M.P. 2008. *Pseudomonads*: major antagonistic endophytic bacteria to suppress bacterial wilt pathogen *Ralstonia solanacearum* in the eggplant (*Solanum melongena* L.). *World Journal of Microbiology and Biotechnology.* 25(1): 47-55.
- [31] Tondje, P.R., Roberts, D.P., Bon, M.C., Widmer, T., Samuels, G.J., Ismaiel, A., Begoude, A.D. Tchana, T., Nyemb- Tshomb, E., Ndoumbe-Nkeng, M., Bateman, R.P., Fontem, D., Hebbar, K.P., 2007. Isolation and identification of mycoparasitic isolates of *Trichoderma asperellum* with potential for suppression of black pod disease of cacao in Cameroon. *Biol. Control* 43, 202-212.
- [32] Deberdt, P., Mfegue, C.V., Tondjé, P.R., Bon, M.C., Cucamp, M., Hurard, C., Begoude, B.A.D., Ndoume-Nkeng, M., Helbar, P.K., Cilas, C., 2008. Impact of environmental factors. Fungicides and biological control on cocoa pod on cocoa pod production dynamics and black pod diseases (*Phytophthora megakarya*) in Cameroon. *Biol. Control* 44, 149-159.
- [33] Pedro Ulises Bautista-Rosales, Montserrat Calderon-Santoyo, Rosalía Servín-Villegas, Norma Angélica Ochoa-Álvarez, Juan Arturo Ragazzo-Sánchez (2013) Action mechanisms of the yeast *Meyerozyma caribbica* for the control of the phytopathogen *Colletotrichum gloeosporioides* in mangoes. *Biological Control* 65,293–301.
- [34] S.S. Ardakani, A. Heydari, N. Khorasani and R. Arjmandi (2010) Development of new bioformulations of *Pseudomonas fluorescens* and evaluation of these products against damping-off of cotton seedlings. *Journal of Plant Pathology.* 92 (1), 83-88.
- [35] Heydari A., Misaghi I.J., 1998. The impact of herbicides on the incidence and development of *Rhizoctonia solani* induced cotton seedling damping-off. *Plant Disease* 82: 110-113.
- [36] Heydari A., Misaghi I.J., 2003. The role of rhizosphere bacteria in herbicide-mediated increase in *Rhizoctonia solani* induced cotton seedling damping-off. *Plant and Soil* 257:391-396.
- [37] Bharathi R., Vivekananthan R., Harish S., Ramanathan A., Samiyappan R., 2004. Rhizobacteria-based bioformulations for the management of fruit rot infection in chilies. *Crop Protection* 23: 835-843.

- [38] Heydari A., Gharedaghi A., 2007. Integrated Pest Management on Cotton in Asia and North Africa. INCANA Press, Tehran, Iran.
- [39] Aeron, A., R.C. Dubey, D.K. Maheshwari, P. Pandey, V.K. Bajpai and S.C. Kang: (2011) Multifarious activity of bioformulated *Pseudomonas fluorescens* PS1 and biocontrol of *Sclerotinia sclerotiorum* in Indian rapeseed (*Brassica campestris* L.). *Eur. J. Pl. Pathol.* 131, 81-93.
- [40] P.P.Jambhulkar and P. Sharma (2014) Development of bioformulation and delivery system of *Pseudomonas fluorescens* against bacterial leaf blight of rice (*Xanthomonas oryzae* pv. *oryzae*). *Journal of Environmental Biology*, 35, 843-849.
- [41] Kosanke JW, Osburn RM, Shuppe GI, Smith RS (1992) Slow rehydration improves the recovery of dried bacterial populations. *Can J Microbiol* 38:520–525.
- [42] Bashan Y (1998) Inoculants of plant growth promoting bacteria use in agriculture. *Biotech Adv* 6:729–770.
- [43] Shah-Smith DA, Burns RG (1997) Shelf-life of a biocontrol *Pseudomonas putida* applied to sugar beet seeds using commercial coating. *Biocontrol Sci Technol* 7:65–74.
- [44] Paul E, Fages J, Blanc P, Goma G, Pareilleux A (1993) Survival of alginate-entrapped cells of *Azospirillum lipoferum* during dehydration and storage in relation to water properties. *Appl Microbiol Biotechnol* 40:34–39.
- [45] Saha AK, Deshpande MV, Kapadnis BP (2001) Studies on survival of *Rhizobium* in the carriers at different temperatures using green fluorescent protein marker. *Curr Sci* 80(5):669–671.
- [46] Duffy BK, De fago G (1999) Environmental factors modulating antibiotic and siderophore biosynthesis by *Pseudomonas fluorescens* biocontrol strains. *Appl Environ Microbiol* 65:2429–2438.
- [47] Bell A, Hubbard JC, Liu L, Davis RM, Subbarao KV (1998) Effects of chitin and chitosan on the incidence and severity of *Fusarium* yellows of celery. *Plant Dis* 82:322–328.
- [48] Kishore GK, Pande S, Podile AR (2005) Biological control of late leaf spot of peanut (*Arachis hypogea* L.) with chitinolytic bacteria. *Phytopathology* 95:1157–1165.
- [49] Raj NS, Deepak SA, Basavaraju P, Shetty HS, Reddy MS, Klopper WJ (2003). Comparative performance of formulations of plant growth promoting rhizobacteria in growth promotion and suppression of downy mildew in pearl millet. *Crop Protection* 22:579-588.
- [50] Chuaboon W, Prathuangwong S (2007). Biological control of cauliflowerer soft rot using bacterial antagonist and its risk assessment. *J. Thai Phytopathol.* 21:63-48.
- [51] Prathuangwong S, Chuaboon W, Kasem S, Hiromitsu N, Suyama K (2007). Formulation development of *Pseudomonas fluorescens* SP007s to control Chinese kale diseases in farming production. Abstract of paper. In: Proceedings of the ISSAAS Int. Cong. Agriculture Is a Business, Dec 12–14, Melaka, 58.
- [52] Commare RR, Nandakumar R, Kandan A, Suresh S, Bharathi M, Raguchander T, Samiyappan R (2002) . *Pseudomonas fluorescens* based bio-formulation for the management of sheath blight disease and leafhopper insect in rice. *Crop Protection* 21:671–677.
- [53] Cook RJ (2002). Advances in plant health management in the twentieth century. *Annu. Rev. Phytopathol.* 38:95-116.
- [54] Bharathi R, Vivekananthan R, Harish S, Ramanathan A, Samiyappan R (2004). Rhizobacteria-based bio-formulations for the management of fruit rot infection in chillies. *Crop Protection* 23:835-843.
- [55] Chaluvvaraju G, Basavaraju P, Shetty NP, Deepak SA, Amruthesh KN, Shetty HS (2004). Effect of some phosphorous based compounds on control of pearl millet downy mildew disease. *Crop Protection* 23:595-600.
- [56] Anitha A, Rabeeth M (2009). Control of fusarium wilt of tomato by bioformulation of *Streptomyces griseus* in green house condition. *Afr. J. Basic Appl. Sci.* 1(1-2):9-14.
- [57] Chen XH, Scholz R, Borriss M, Junge H, MÖgeI G, Kunz S, Borriss R (2009). Difficidin and bacilysin produced by plant-associated *Bacillus amyloliquefaciens* are efficient in controlling fire blight disease. *J. Biotech.* 140(1-2):38-44.
- [58] Ardakani SS, Heydari A, Khorasani N, Arjmandi R (2010). Development of new bioformulations of *Pseudomonas fluorescens* and evaluation of these products against damping-off cotton seedlings. *J. Plant Pathol.* 92(1):83-88.
- [59] Ardakani SS, Heydari A, Tayebi L, Cheraghi M (2011). Evaluation of efficacy of new bioformulations on promotion of cotton seedlings. *Environ. Sci. Technol.* 6:361-364.
- [60] Haggag M, Wafaa SS (2012). Development and production of formulations of PGPR cells for control of leather fruit rot disease of strawberry. *Am. J. Sci. Res.* 67:16-22.
- [61] Schisler DA, Slininger PJ, Behle RW, Jackson MA (2004). Formulation of *Bacillus* spp. for biological control of plant diseases. *Phytopathology* 94 (11):1267-1271.
- [62] Sharathchandra RG, Raj NS, Shetty NP, Amruthesh KN, Shetty SH (2004). A Chitosan formulation Elexat induces downy mildew disease resistance and growth promotion in pearl millet. *Crop Protection* 23: 881-888.
- [63] Amran M (2006). B iomass production and formulation *Bacillus subtilis* for biological control. *Indonesian J. Agri. Sci.* 7(2):51-56.
- [64] Pushpalatha HG, Mythrashree SR, Shetty R, Geetha NP, Sharathchandra RG, Amruthesh KN, ShettySH (2007). Ability of vitamins to induce downy mildew disease resistance and growth promotion in pearl millet. *Crop Protection* 26:1674-1681.
- [65] Preecha C, Prathuangwong S (2009). Development of *Bacillus amyloliquefaciens* KPS46 formulation for control of soybean disease. Abstract of paper. In: Proceedings of the ISSAAS Int.Conf., Feb 23-27, Bangkok, 222.
- [66] Omer MA (2010). Bioformulations of *Bacillus* spores for using as Biofertilizer. *Life Sci. J.* 7:4.
- [67] Senthilraja G, Anand T, Durairaj C, Raguchander T, Samiyappan R (2010). Chitin-based bioformulation of *Beauveria bassiana* and *Pseudomonas fluorescens* for improved control of leafminer and collar rot in groundnut. *Crop Protection* 29:1003-1010.
- [68] Siripornvisal S, Trilux S (2011). Effect of a bioformulation containing *Bacillus subtilis* BCB3-19 on early growth of hongtae pak choi. *Agri. Sci. J.* 42(2):293-296.
- [69] Agrawal Pushpa, Pandey Subhash C. and Manjunatha Reddy A.H. (2014) Development of liquid formulation for the Dual purpose of crop protection and Production *Journal of Environmental Research and Development.* 8(3):378-383.
- [70] Chakraborty U, Chakraborty B N, Chakraborty AP, Sunar K and Dey PL (2013) Plant growth promoting rhizobacteria mediated improvement of health status of tea plants. *Indian Journal of Biotechnology.* 12 (1):20-31.
- [71] Bashan Y, Inoculants of plant growth promoting rhizobacteria for use in agriculture, *Biotechnol Adv*, 16 (1998) 729-770.
- [72] El-Hassan S A & Gowen S R, Formulation and delivery of the bacterial antagonist *Bacillus subtilis* for management of lentil vascular wilt caused by *Fusarium oxysporum* f. sp. *lentis*, *J Phytopathol*, 154 (2006) 148-155.
- [73] Trivedi P, Pandey A & Palni L M S, Carrier-based preparations of plant growth promoting bacterial inoculants suitable for use in cooler regions, *World J Microbiol Biotechnol*, 21 (2005) 941-945.
- [74] Suriyaprabha Rangaraj, Karunakaran Gopalu, Prabhu Muthusamy, Yuvakkumar Rathinam, Rajendran Venkatachalam and Kannan Narayanasamy (2014). Augmented biocontrol action of silica nanoparticles and

- Pseudomonas fluorescens* bioformulant in maize (*Zea mays* L.). RSC Adv.4, 8461–8465.
- [75] Smith RS (1992) Legume inoculant formulation and application. *Can J Microbiol* .38, 485–492.
- [76] Rzewnicki P (2000) Ohio organic producers: final survey results. Online. Ohio State University Extension, College of Food Agricultural and Environmental Sciences, Bulletin, Special Circular, 174.
- [77] Validov S, Kamilova F, Qi S, Stephen D, Wang JJ, Makarova N, Lugtenberg B (2007) Selection of bacteria able to control *Fusarium oxysporum* f. sp. *Radicus-lycopersici* in stone substrate. *J Appl Microbiol* 102:461–471.
- [78] Validov SZ, Kamilova F, Lugtenberg BJJ (2009) *Pseudomonas putida* strain PCL1760 controls tomato foot and root rot in stonewool under industrial conditions in a certified greenhouse. *Biol Control* 48:6–11.
- [79] Fages J (1992) An industrial view of *Azospirillum* inoculants formulation and application technology. *Symbiosis* 13:15–26.
- [80] Garcí a de Castro A, Lapinski J, Tunnacliffe A (2000) Anhydrobiotic engineering. *Nat Biotechnol* 18:473.
- [81] Bharat C. Nath, L.C. Bora, L. Katakí, K. Talukdar, P. Sharma, J. Dutta and P. Khan (2016). Plant Growth Promoting Microbes, their Compatibility Analysis and Utility in Biointensive Management of Bacterial Wilt of Tomato. *Int.J.Curr.Microbiol.App.Sci* 5, 1007-1016.
- [82] Cook, R.J., and Baker, K.F. (1983). *The Nature and Practice of Biological Control of Plant Pathogens*. (St. Paul, MN: APS Press 281 pp).
- [83] Francisco D.H., Angelica M.P., Gabriel M., Melchor C.S., Raul R., Cristobal N., Francisco C.R. (2011). In vitro antagonist action of *Trichoderma* strains against *Sclerotium sclerotiorum* and *Sclerotium cepivorum*. *American Journal of Agriculture Biology Science* 6 (3), 410–417.
- [84] Govind Gupta, Shailendra Singh Parihar, Narendra Kumar Ahirwar, Sunil Kumar Snehi and Vinod Singh (2015). Plant Growth Promoting Rhizobacteria (PGPR): Current and Future Prospects for Development of Sustainable Agriculture. *J. Microb. Biochem. Technol.* 7, 96-102.
- [85] Heydari A, Pessarakli M. (2010). A review on biological control of fungal plant pathogens using microbial antagonists. *J Biol Sci* 10, 272-290.
- [86] Kakvan N, Heydari A, Zamanizadeh HR, Rezaee S, Nraghi L.(2013). Development of new bioformulations using *Trichoderma* and *Talaromyces* fungal antagonists for biological control of sugar beet damping-off disease. *Crop Prot* 53, 80-84.
- [87] Leta A, Selvaraj TH.(2013). Evaluation of Arbuscular mycorrhizal fungi and *Trichoderma* species for the control of onion white rot (*Sclerotium cepivorum* Berk). *Plant Patho Microbiol* 14, 1-6.
- [88] Malleswari Damam, Bagyanarayana Gaddam and Rana Kausar (2015). Bio-Management of Root-Rot Disease Caused by *Macrophomina phaseolina* in *Coleus forskohlii*. *IJPPR*. 7, 347-352.
- [89] Metcalf DA, Dennis JJC, Wilson CR. (2004). Effect of inoculum density of *Sclerotium cepivorum* on the ability of *Trichoderma koningii* to suppress white rot of onion. *Plant Dis* 88, 287-291.
- [90] Naraghi L, Heydari A, Rezaee S, Razavi M. (2013). Study on some antagonistic mechanisms of *Talaromyces flavus* against *Verticillium dahliae* and *Verticillium albo-atrum*, the causal agents of wilt disease in several important crops. *Biocont Plant Prot* 1,13-28.
- [91] Razak Mahdizadehnaraghi, Asghar Heydari, Hamid Reza Zamanizadeh, Saeed Rezaee, Jafar Nikan (2015). Promotion of Garlic Growth Characteristics Using Bioformulations Developed Based on Antagonistic Fungi. *Intl. J. Agri. Crop Sci.* 8, 654-658.
- [92] Suárez-Estrella, F., Vargas-García, C., López, M.J., Capel, C., Moreno, J., 2007. Antagonistic activity of bacteria and fungi from horticultural compost against *Fusarium oxysporum* f. sp. *melonis*. *Crop Prot.*, 46–53.
- [93] El-Hasan, A., Walker, F., Buchenauer, H., 2006. *Trichoderma harzianum* and its metabolite 6-pentyl-alpha-pyrone suppress fusaric acid produced by *Fusarium moniliforme*. *J. Phytopathol.* 156, 79–87.
- [94] Gava, C.A.T., Menezes, M.E.L., 2012. Efficiency of *Trichoderma* spp isolates on the control of soil-borne pathogens yellow melon in field conditions. *Rev. Ciência Agronômica* 43, 633–640.
- [95] Woo, S.L., Ruocco, M., Vinale, F., Nigro, M., Marra, R., Lombardi, N., Pascale, A., Lanzuise, S., Manganiello, G., Lorito, M., 2014. *Trichoderma*-based products and their widespread use in agriculture. *Open Mycol. J.* 8, 71–126.
- [96] Shahid, M., Srivastava, M., Pandey, S., Singh, A., Kumar, V., Srivastava, Y., 2014. Biocontrol mechanisms by *Trichoderma* through genomics and proteomics analysis: a review. *Afr. J. Microbiol. Res.* 8, 3064–3069.
- [97] Harman, G.E., Howell, C.R., Viterbo, A., Chet, I., Lorito, M., 2004. *Trichoderma* species opportunistic, avirulent plant symbionts. *Nat. Rev. Microbiol.* 2, 43–56.
- [98] Reino, J.L., Guerrero, R.F., Hernández-Galán, R., Collado, I.G., 2008. Secondary metabolites from species of the biocontrol agent *Trichoderma*. *Phytochem. Rev.* 7, 89–123.
- [99] Fravel, D.R., 2005. Commercialization and implementation of biocontrol. *Annu. Rev. Phytopathol.* 43, 337–359.
- [100] Martínez-Medina, A., Pascual, J.A., Pérez-Alfocea, L.F., Roldán, A., 2010. *Trichoderma harzianum* and *Glomus intraradices* modify the hormone disruption induced by *Fusarium oxysporum* infection in melon plants. *Phytopathology* 100, 682–688.
- [101] Yedidia, I., Shores, M., Kerem, Benhamou, N., Kapulnik, Y., Chet, I., 2003. Concomitant induction of systemic resistance to *Pseudomonas syringae* pv. *lachrymans* in cucumber by *Trichoderma asperellum* (t-203) and accumulation of phytoalexins. *Appl. Environ. Microbiol.* 69, 7343–7353.